

PHYTOECDYSTEROIDS FROM THE *Silene* GENUS

N. Z. Mamadalieva,¹ L. N. Zibareva,² R. Lafont,³
L. Dainan,⁴ and Z. Saatov¹

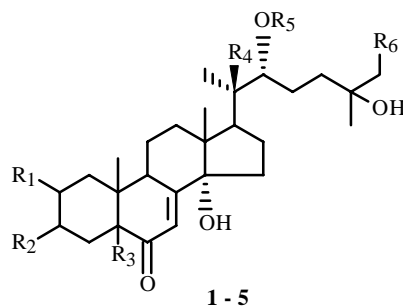
UDC 581.3.82:547.926

Research results on the isolation and identification of ecdysteroids from plants of the *Silene* L. genus were presented.

Key words: phytoecdysteroids, distribution, isolation, *Silene* genus.

The genus *Silene* L. is a good source of new ecdysteroid analogs [1-5]. Species of this genus contain 59% of ecdysteroids. Ecdysteroids have been found by us in 115 species of *Silene* including in 75 species for the first time [6]. Of these, 26 species had a high content of them. Until now the chemical composition of these compounds has been determined in 23 *Silene* species including 12 investigated by us.

We established for the first time the ecdysteroid profile of the following species: *Silene antirrhina*, *S. chlolirifolia*, *S. cretica*, *S. disticha*, *S. echinata*, *S. italica*, *S. linicola*, *s. portensis*, *S. pseudotites*, *S. radicata*, *S. regia*, *S. viridiflora* [4, 5, 7] using a developed detection method in seeds [8]. These data were subsequently confirmed by radioimmunoassay (Table 1). The studied species possess agonistic (ecdysteroid) activity but not one of them exhibited antagonistic activity. This confirms that they contain phytoecdysteroids.



1 - 5

- 1: R₁ = R₃ = R₅ = H, R₂ = R₄ = R₆ = OH
2: R₁ = R₃ = R₄ = R₆ = H, R₂ = OH, R₅ = β-D-Glc
3: R₁ = R₅ = R₆ = H, R₂ = β-D-Glc, R₃ = R₄ = OH
4: R₁ = R₄ = R₆ = OH, R₂ = OAc, R₃ = H, R₅ = Ac
5: R₁ = OAc, R₂ = R₄ = R₆ = OH, R₃ = H, R₅ = OAc

1) S. Yu. Yunusov Institute of the Chemistry of Plant Substances, Academy of Sciences, Republic of Uzbekistan, Tashkent, fax (99871) 120 64 75; 2) Tomsk University Siberian Botanical Garden, Tomsk; 3) Universite Pierre et Marie Curie, Laboratoire d'Endocrinologie Moléculaire et Evolution, France; 4) Department of Biological Sciences, Hatherly Laboratories, Prince of Wales Road, Exeter, UK. Translated from *Khimiya Prirodnykh Soedinenii*, No. 6, pp. 472-475, November-December, 2004. Original article submitted June 21, 2004.

TABLE 1. Analysis of Caryophyllaceae Plant Extracts for Ecdysteroid Content

Plant species	Growth phase	Plant part	CSP analysis	RIA DBL-1	Biotest for agonists
<i>S. antirrhina</i>	Fr	Fruit,	0.5	969	+++
		seeds	0.1	87.2	++
<i>S. echinata</i>	Fl	Flowers,	0.4	5972	+++
	Fr	seeds	0.1	101.4	++
<i>S. viridiflora</i>	B	Buds,	1.6	6582	++++
	Fr	seeds	0.2	659	++++
<i>S. frivaldszkyana</i>	Fl	Flowers,	6.2	27592	++++
	Fr	seeds	0.6	113.2	++++
<i>S. portensis</i>	Fl	Flowers,	1.4	15312	C+++
	Fr	seeds	0.2	86.2	++
<i>S. linicola</i>	Fl	A. part,	0.5	2000	+++
	Fr	seeds	0.6	1485	+++(+)-
<i>S. radicata</i>	Fl	Flowers	1.3	11393	++++
<i>S. pseudotites</i>	E	Leaves	0.6	27188	+++
<i>S. italica</i>	Fl	Flowers	1.3	12205	++++
<i>S. cretica</i>	Fr	Fruit,	0.4	1920	+++
		seeds	0.3	49.1	++
<i>S. disticha</i>	Fl	Leaves,	0.4	2555	C++
	Fr	seeds	0.3	109.5	++
<i>S. chlorifolia</i>	E	Leaves	0.8	3020	+++

B, budding; Fl, flowering; Fr, fruiting; E, end of vegetation; A. part, aerial part. CSP, chromatophotometric method, % of dry material. RIA, radioimmunoassay, μg ecdysone equivalent/g. C, cytotoxic; -, inactive; +, undiluted extract active; (+), weakly active; ++, +++, +++++, active at 10-, 100-, and 1000-fold, respectively, dilution.

The main ecdysteroid components were identified in seven species (*S. antirrhina*, *S. chlorifolia*, *S. cretica*, *S. disticha*, *S. echinata*, *S. italica*, *S. portensis*) by HPLC using the UV spectra of the peaks. The absorption maximum is due to the presence of a conjugated ketone and occurs near 242-250 nm. Semipreparative HPLC over a C_{18} column was used to concentrate the pure compounds. The retention times of the resulting peaks compared with those of polypodine B (P_B), 20-hydroxyecdysone (20E), 25R-inocosterone, 25S-inocosterone, 2-deoxy-20-hydroxyecdysone, and ecdysone and co-injection with the proposed ecdysteroid standards enabled the compounds to be identified on two analytical reversed-phase C_{18} and C_6 columns (Table 2). The major components in each *Silene* species are 20E and P_B . Ecdysteroids 1-5 were isolated for the first time. Glycosides are typically found for the *Silene* genus. For example, the series of glycosides sileneoside A, B, C, D, and E was found in *S. brachiuca* [9-13], *S. nutans*, *S. scabrifolia*, *S. supina*, *S. viridiflora* [14, 15]; in *S. otites*, 20E 25-glycoside [3]. However, 2-deoxyecdysone 22 β -D-glycoside and 2-deoxypolypodine B 3-O- β -D-glycoside were isolated for the first time. The presence in plants of the ecdysteroid glycosides indicates that they play an active role in metabolic processes [1].

EXPERIMENTAL

Extracts were purified using Sep-Pac Vac 35-cc cartridges. A Gilson Model 811 HPLC chromatograph combined with a Gilson 160 detector and the Gilson Unipoint computer program were used for analysis. HPLC separation of the extracts was performed over Spherisorb 5 ODS-2 analytical columns with C_{18} and C_6 (particle size 5 mm, 150 \times 4.6 mm, Jones Chromatography).

TABLE 2. Phytoecdysteroids Isolated from *Silene* L. Species

Ecdysteroid	Formula	Yield, %	Plant species
20-Hydroxyecdysone	$C_{27}H_{44}O_7$	0.071	<i>S. pseudotites</i>
		0.367	<i>S. linicola</i>
		0.042	<i>S. radicata</i>
		0.302	<i>S. regia</i>
		0.350	<i>S. viridiflora</i>
Polipodine B	$C_{27}H_{44}O_8$	0.006	<i>S. pseudotites</i>
		0.073	<i>S. linicola</i>
		0.007	<i>S. radicata</i>
		0.021	<i>S. regia</i>
		0.250	<i>S. viridiflora</i>
25S-Inocosterone	$C_{27}H_{44}O_7$	Tr.	<i>S. pseudotites</i>
		0.012	<i>S. regia</i>
Ecdysone	$C_{27}H_{44}O_6$	0.002	<i>S. pseudotites</i>
		0.001	<i>S. linicola</i>
2-Deoxyecdysone	$C_{27}H_{44}O_5$	0.165	<i>S. pseudotites</i>
2-Deoxy-20-hydroxyecdysone	$C_{27}H_{44}O_6$	0.106	<i>S. pseudotites</i>
		0.017	<i>S. linicola</i>
		0.200	<i>S. viridiflora</i>
Ponasterone A	$C_{27}H_{44}O_6$	Tr.	**
Sidisterone	$C_{24}H_{32}O_6$	0.001	<i>S. pseudotites</i>
2-Deoxyintegristerone A	$C_{27}H_{44}O_7$	0.001	<i>S. pseudotites</i>
(5 α)-2-Deoxyintegristerone A	$C_{27}H_{44}O_7$	0.001	<i>S. pseudotites</i>
2-Deoxy-21-hydroxyecdysone	$C_{27}H_{44}O_6$	0.013	<i>S. pseudotites</i>
Viticosterone E	$C_{29}H_{46}O_8$	0.001	<i>S. linicola</i>
Turkesterone	$C_{27}H_{44}O_8$	0.001	<i>S. linicola</i>
Integristerone A	$C_{27}H_{44}O_8$	0.002	<i>S. linicola</i>
		0.200	<i>S. viridiflora</i>
Sileneoside A	$C_{33}H_{54}O_{12}$	0.080	<i>S. viridiflora</i>
Sileneoside D	$C_{33}H_{54}O_{12}$	0.100	<i>S. viridiflora</i>
26-Hydroxypolipodine B	$C_{27}H_{44}O_9$	0.035	<i>S. viridiflora</i>
2-Deoxy-20,26-dihydroxyecdysone (1)*	$C_{27}H_{44}O_7$	0.003	<i>S. pseudotites</i>
2-Deoxyecdysone 22 β -D-glycoside (2)*	$C_{33}H_{54}O_{10}$	0.003	<i>S. pseudotites</i>
2-Deoxypolipodine B 3 β -D-glycoside (3)*	$C_{33}H_{54}O_{12}$	0.003	<i>S. pseudotites</i>
20,26-Dihydroxyecdysone 3,22-diacetate (4)*	$C_{31}H_{48}O_{10}$		<i>S. viridiflora</i>
20,26-Dihydroxyecdysone 2,22-diacetate (5)*	$C_{31}H_{48}O_{10}$		<i>S. viridiflora</i>

*Ecdysteroids isolated for the first time, **detected in most species; Tr.: traces.

Radioimmunoassay and biotest B_{II} for the presence and determination of ecdysteroid content in the plants were carried out at Exeter University (Exeter, Great Britain). IR spectra were recorded on a Perkin—Elmer 2000 FT-spectrometer. NMR spectra were recorded on JNM-4H-100, Bruker WP 200 SY, Bruker Avance DRX400, and Bruker AMX500 instruments at 27°C using standard Bruker microprograms. The solvents were C₅D₅N, CD₃OD, and D₂O with hexamethyldisiloxane (HMDS), tetramethylsilane (TMS), and sodium [2,2,3,3-²H₄]-3-(trimethylsilyl)propionate internal standards.

TABLE 3. ^1H and ^{13}C NMR Chemical Shifts of 20,26-Dihydroxyecdysone 3,22-Diacetate (**4**) and 20,26-Dihydroxyecdysone 2,22-Diacetate (**5**) (D_2O , δ , ppm, J/Hz)

C atom	^{13}C (4)	^1H (4)	^{13}C (5)	^1H (5)
1	37.7	1.48, 1.99	37.7	1.53, 1.96
2	-	4.13 (m, $w_{1/2} = 22$)	-	5.08 (m, $w_{1/2} = 22$)
3	72.3	5.17 (m, $w_{1/2} = 8$)	73.0	4.22 (m, $w_{1/2} = 8$)
4	-	1.78, 1.85	-	1.80, 1.83
5	51.9	2.35 (dd, $J = 4.2, 13.5$)	51.5	2.41 (dd, 5.13)
6	-	-	-	-
7	122.4	5.98 (d, $J = 2.5$)	122.3	5.99 (d, $J = 2.5$)
8	-	-	-	-
9	35.0	3.11 (m, $w_{1/2} = 24$)	35.1	3.18 (m, $w_{1/2} = 24$)
10	39.1	-	39.5	-
11	-	1.74, 1.86	-	1.73, 1.86
12	32.0	1.76, 1.96	32.3	1.76, 1.96
13	48.5	-	48.7	-
14	86.3	-	86.6	-
15	31.3	1.67, 2.06	31.6	1.67, 2.06
16	20.7	1.88, 1.76	21.4	1.88, 1.76
17	50.6	2.31 (t, $J = 9.5$)	50.6	2.31 (t, $J = 9.5$)
18	18.1	0.86 s	18.2	0.86 s
19	24.2	1.02 s	24.1	1.02 s
20	78.4	-	78.4	-
21	21.3	1.34 s	21.5	1.34 s
22	82.7	4.85 (dd, $J = 10.5, 2$)	81.4	4.85 (dd, $J = 10.5, 2$)
23	-	1.56, 1.76	-	1.56, 1.76
24	35.9	1.75, 1.46	35.9	1.75, 1.46
25	74.4	-	74.5	-
26	69.9	3.42 s	69.8	3.42 s
27	23.2	1.15 s	23.1	1.15 s
2- CH_3CO	-	-	22.1/175.0	2.127 s
3- CH_3CO	21.7/175.1	2.176 s	-	-
22- CH_3CO	21.7/176.0	2.165 s	21.9/176.1	2.167 s

-, not determined

Mass spectra were recorded on Finnigan MAT-8200 and JEOL JMS-700 (desorption-chemical ionization) mass spectrometers using ammonia as a reagent. Certain mass spectra were obtained on a triple quadrupole (Quattro triple quadrupole) VG mass spectrometer with liquid secondary-ion ionization (LSIMS) using a Cs^+ beam and a glycerine matrix.

2-Deoxy-20,26-dihydroxyecdysone (1), $\text{C}_{27}\text{H}_{44}\text{O}_7$, M.W. 480, UV spectrum (EtOH, λ_{max} , nm, log ϵ): 242 (4.01).

HPLC: t_{R} 23.7 [Zorbax-Sil, $\text{CH}_2\text{Cl}_2:(\text{CH}_3)_2\text{CHOH}:\text{H}_2\text{O}$, 125:40:3]. Mass spectrum (CI, NH_3) (m/z , I_{rel} , %): 498 (81) $[\text{M} + \text{H} + \text{NH}_3]^+$, 481 (54) $[\text{M} + \text{H}]^+$, 480 (100) $[\text{M}]^+$, 463 (10.5) $[\text{M} + \text{H} - \text{H}_2\text{O}]^+$, 445 (4) $[\text{M} + \text{H} - 2\text{H}_2\text{O}]^+$.

PMR spectrum (D_2O , δ , ppm, J/Hz): 0.86 (3H, s, C_{18} -Me), 0.98 (3H, br.s, $w_{1/2} = 4$, C_{19} -Me), 1.16 (3H, s, C_{27} -Me), 1.24 (3H, s, C_{21} -Me), 2.33 (t, $J = 9.7$, H-17), 2.39 (br.dd, $J = 12.5$, $J = 3.4$, H-5), 3.15 (br.s, $w_{1/2} = 25$, H-9), 3.44 (d, $J = 10.5$, H-22), 3.45 [s, $\text{CH}_2\text{OH}(26)$], 5.96 (d, H-7, $J = 2.2$), 4.10 (br.s, $w_{1/2} = 20$, H-3).

2-Deoxyecdysone 22 β -D-glycoside (2), $\text{C}_{33}\text{H}_{54}\text{O}_{10}$, M.W. 610, HPLC: t_{R} 23.5 [Zorbax-Sil, $\text{CH}_2\text{Cl}_2:(\text{CH}_3)_2\text{CHOH}:\text{H}_2\text{O}$, 125:40:3].

Mass spectrum (CI, NH_3) (m/z , I_{rel} , %): 628 (100) $[\text{M} + \text{H} + \text{NH}_3]^+$, 611 (18) $[\text{M} + \text{H}]^+$, 610 (41) $[\text{M}]^+$, 593 (7) $[\text{M} + \text{H} - \text{H}_2\text{O}]^+$.

PMR spectrum (D₂O, δ , ppm, J/Hz): 0.74 (3H, s, C₁₈-Me), 0.956 (3H, d, J = 6.8, C₂₁-Me), 0.990 (3H, br.s, w_{1/2} = 4, C₁₉-Me), 1.238 (3H, s, C₂₆-Me), 1.243 (3H, s, C₂₇-Me), 2.18 (m, H-17), 2.41 (dd, J = 12.3, J = 3.2, H-5), 3.14 (br.m, w_{1/2} = 25, H-9), 3.73 (br.d, J = 10.5, H-22), 4.10 (br.s, w_{1/2} = 23, H-3), 5.97 (d, J = 2.1, H-7), 4.53 (d, J = 8, H-1'), 3.29 (dd, J = 9.1, J = 8.1, H-2'), 3.48 (t, J = 9.3, H-3'), 3.38 (t, J = 9.3, H-4'), 3.45 (m, H-5'), 3.70 (dd, J = 12.4, J = 6.2, H-6'), 3.89 (dd, J = 12.3, J = 2.3, H-6').

¹³C NMR (C₅D₅N, δ , ppm): 101.8 (C-1).

2-Deoxypolipodine B 3 β -D-glycoside (3), C₃₃H₅₄O₁₂, M.W. 642. HPLC: t_R 38.0 [Zorbax-Sil, CH₂Cl₂:(CH₃)₂CHOH:H₂O, 125:40:3].

Mass spectrum (CI, NH₃) (*m/z*, I_{rel}, %): 480 (9) [M - sugar]⁺, 463 (40) [M + H - sugar - H₂O]⁺, 180 (46) [sugar]⁺.

PMR spectrum (D₂O, δ , ppm, J/Hz): 0.875 (3H, s, C₁₈-Me), 0.905 (3H, s, C₁₉-Me), 1.221 (3H, s, C₂₆-Me), 1.233 (3H, s, C₂₁-Me), 1.233 (3H, s, C₂₇-Me), 3.44 (d, J = 10.5, H-22), 2.32 (t, J = 9.3, H-17), 3.243 (m, H-9), 4.31 (br.s, w_{1/2} = 12, H-3), 5.99 (d, J = 2.1, H-7), 4.56 (d, J = 8, H-1'), 3.30 (dd, J = 9, J = 8, H-2'), 3.51 (t, J = 9.3, H-3'), 3.39 (t, J = 9.3, H-4'), 3.45 (m, H-5'), 3.71 (dd, J = 12.4, J = 5.9, H-6'), 3.90 (dd, J = 12.3, J = 2, H-6').

REFERENCES

1. N. K. Abubakirov, *Khim. Prir. Soedin.*, 685 (1981).
2. Z. Saatov, Author's Abstract of a Doctoral Dissertation in Chemical Sciences, Tashkent (1993).
3. J.-P. Girault, M. Bathori, E. Varga, K. Szendrei, and R. Lafont, *J. Nat. Prod.*, **53**, 279 (1990).
4. L. N. Zibareva, *Rastit. Resur.*, **35**, No. 1, 79 (1999).
5. L. Zibareva, *Arch. Insect. Biochem. Physiol.*, **43**, 1 (2000).
6. L. N. Zibareva, Author's Abstract of a Doctoral Dissertation in Chemical Sciences, Novosibirsk (2003).
7. L. N. Zibareva, *Rastit. Resur.*, 89 (1997).
8. L. N. Zibareva, V. I. Eremina, and P. V. Zibarev, Pat. No. 2,082,168; MKI 6 G 01 N 33/50, 30/02, 30/90. No. 94002629/13; Filed Jan. 26, 1994; Issued June 20, 1997; *Byull.* No. 17, Priority Jan. 26, 1994.
9. Z. Saatov, M. B. Gorovits, N. D. Abdullaev, B. Z. Usmanov, and N. K. Abubakirov, *Khim. Prir. Soedin.*, 738 (1981).
10. Z. Saatov, M. B. Gorovits, N. D. Abdullaev, B. Z. Usmanov, and N. K. Abubakirov, *Khim. Prir. Soedin.*, 611 (1982).
11. Z. Saatov, M. B. Gorovits, N. D. Abdullaev, B. Z. Usmanov, and N. K. Abubakirov, *Khim. Prir. Soedin.*, 211 (1982).
12. Z. Saatov, N. D. Abdullaev, M. B. Gorovits, and N. K. Abubakirov, *Khim. Prir. Soedin.*, 741 (1984).
13. Z. Saatov, N. D. Abdullaev, M. B. Gorovits, and N. K. Abubakirov, *Khim. Prir. Soedin.*, 323 (1986).
14. N. Sh. Ramazanov, S. A. Sultanov, Z. Saatov, and A. M. Nigmatullaev, *Khim. Prir. Soedin.*, 718 (1997).
15. N. Z. Mamadalieva, L. N. Zibareva, N. Edvard-Todeschi, J.-P. Girault, A. Maria, N. Sh. Ramazanov, Z. Saatov, and R. Lafont, *Collect. Czech. Chem. Commun.*, **69**, 1675 (2004).